Distinct roles for mTOR in generating versus sustaining humoral immunity

Derek D. Jones¹, Brian T. Gaudette¹, Joel R. Wilmore¹, Irene Chernova¹, Brendan M. Weiss², and David Allman¹

¹Department of Pathology and Laboratory Medicine and the ²Department of Medicine, Perelman School of Medicine at the University of Pennsylvania, Philadelphia, PA 19104



Supplemental Figure 1. Naïve B cell subsets are unaffected by mTOR inhibition. (A) Absolute numbers of B cells in the BM or spleen were evaluated for mice of the indicated genotype, following 3 administrations of tamoxifen to induce Raptor deletion. B cell subsets were profiled by analysis of 2.5×10^6 events by flow cytometry. f/f; Tam-cre (n=4), f/+; Tam-cre (n=3), +/+; Tam-cre (n=3), f/f; no cre (n=4). No significant differences were identified between any genotypes. Pre-B (B220⁺ CD43⁻ AA4⁺ IgM⁻); immature (B220⁺ CD43⁻ AA4⁺ IgM⁺); mature (B220⁺ CD43⁻ AA4⁻ IgM⁺); FO (follicular, AA4⁻ CD21^{int} CD23⁺ IgM^{low}); MZ, (marginal zone, AA4⁻ CD21^{high} CD23⁻ IgM^{high}); MZP, (marginal zone precursor, AA4⁻ CD21^{high} CD23⁺ IgM^{high}); T1, transitional (AA4⁺ IgM⁺ CD23⁻); T2 (AA4⁺ IgM⁺ CD23⁻), T3 (AA4⁺ IgM^{int} CD23⁻). (B) C57BL/6 mice were treated 3 times with rapamycin (20mg/kg), Torin 1 (10mg/kg), or everolimus (10mg/kg), followed by quantitation of BM and spleen B cell subsets as in (A). Four mice were used in each group. * p<0.05 as determined using a Kruskal-Wallis test with Dunn's multiple comparison test. Significant differences were only found between control and Torin1-treated mice; no other pairwise comparisons reached significance.



Supplemental Figure 2. TORC1 signaling is dispensable for memory B cell maintenance. (A) Mice of the indicated genotype were immunized, and 45 days later splenocytes were stained with the indicated antibodies and NP-allophycocyanin to detect NP-binding cells. Plots were pre-gated for viable IgD⁻ Dump⁻ CD19⁺ CD38⁺ lymphocytes. (B) Absolute numbers per spleen of NP-binding CD73⁺ PDL2⁺ cells. Dots represent values for individual mice, and horizontal lines indicate the means. No significant differences were detected between any genotypes. Data are representative of 2 independent experiments.



Supplemental Figure 3. GC centroblasts exhibit enhanced sensitivity to rapamycin. (A) Splenocytes from 33-week old NZB/W mice either untreated (control) or treated with twice weekly with rapamycin for 6 weeks were stained with the indicated reagents. Representative plots from 3 independent experiments using 4-6 mice per group. (B) Splenocytes from C57BL/6 mice immunized with NP-C γ G 17 days previously were stained with the indicated antibodies and NP-allophycocyanin. Rapamycin-treated mice were administered inhibitor on days 12, 14, and 16 post-immunization. The phenotype of NP-binding GC B cells was further evaluated for light zone centrocytes (CD86⁺ CXCR4⁻) or dark zone centroblasts (CD86⁻ CXCR4⁺). (C) Absolute numbers of NP-binding CD38⁺ PNA⁺ B cells were determined for the mice depicted in (B, *middle plots*). Dots represent values for individual mice, and horizontal lines indicate the means. * p<0.0001 as determined by Student's t-test comparing control and treated mice. (D) *Top*, absolute numbers of light zone (LZ) centrocytes and dark zone (DZ) centroblasts were evaluated in control and rapamycin-treated mice. *Bottom*, average frequency of cells within the LZ or DZ in control (n=4) and rapamycin-treated (n=5) mice; bars indicate the SEM. * p<0.001 as determined by Student's t-test comparing control and treated mice. Representative of 2 independent experiments.